

Biomechanical devices for bone tissue synthesis and regeneration

PhD Course in **Ingegneria**
XL° cycle

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Rationale and objective

Why bone regeneration is still a challenge

- **critical-size** bone defects exceed the body's healing capacity
- bone repair requires both **mechanical** stability and a favorable **biological** environment



Objective: to investigate mechanical and biological strategies for bone regeneration, with a focus on their potential complementarity

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Additive Manufacturing

- complex geometries and patient-specific implants
- lattice structures → trabecular bone porosity and stiffness
- widely used Ti6Al4V alloy
- numerical validation

Regenerative Medicine

- regeneration depends on local biological environment
- biomaterials + cells + biochemical cues
- hydrogels mimicking ECM
- collagen and GelMA

FDM gyroid PEKK lattices

Mechanical characterization for orthopedic applications

PEKK

- mechanical properties ~ bone
- imaging compatibility
- more suitable for AM

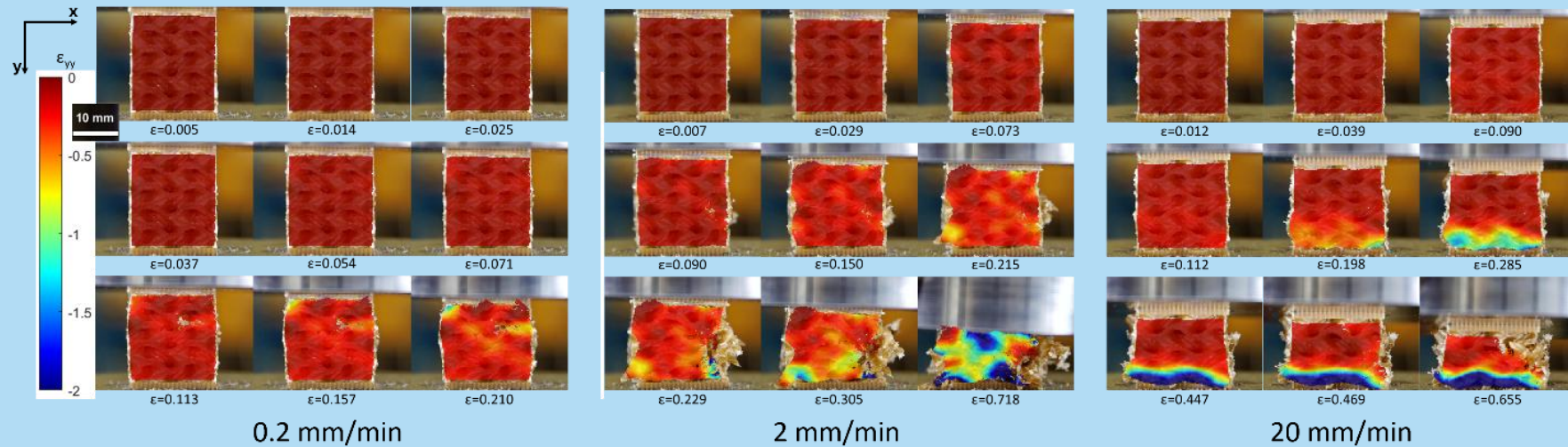


TPMS gyroid structure

- interconnected **porosity**
- surface area for cell attachment (**osseointegration**)
- mechanical tuning through **relative density**

Compression tests

- 300 N **pre-load** and up to large deformations to study onset and **evolution of collapse**
- full-field deformation analysis through Digital Image Correlation (**DIC**)



- constant deformation
- no macroscopic fracture zones

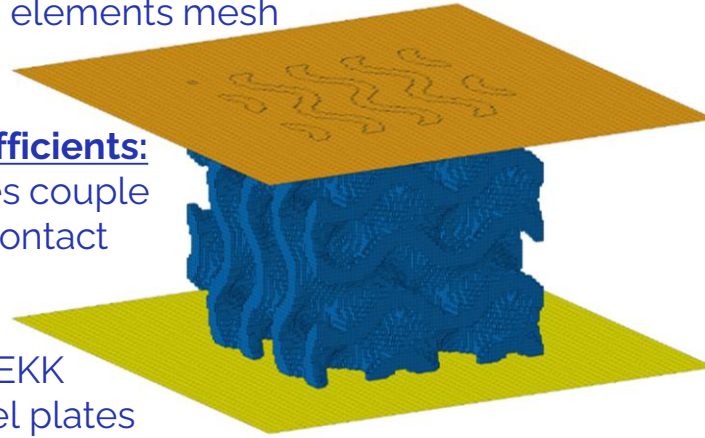
- earlier cell collapse
- diagonal shear bands
→ bending-dominated

- deformation in lower layers
- brittle-like collapse
- higher measured peak forces

FDM gyroid PEKK lattices

Numerical modelling and FEM validation for 20 mm/min

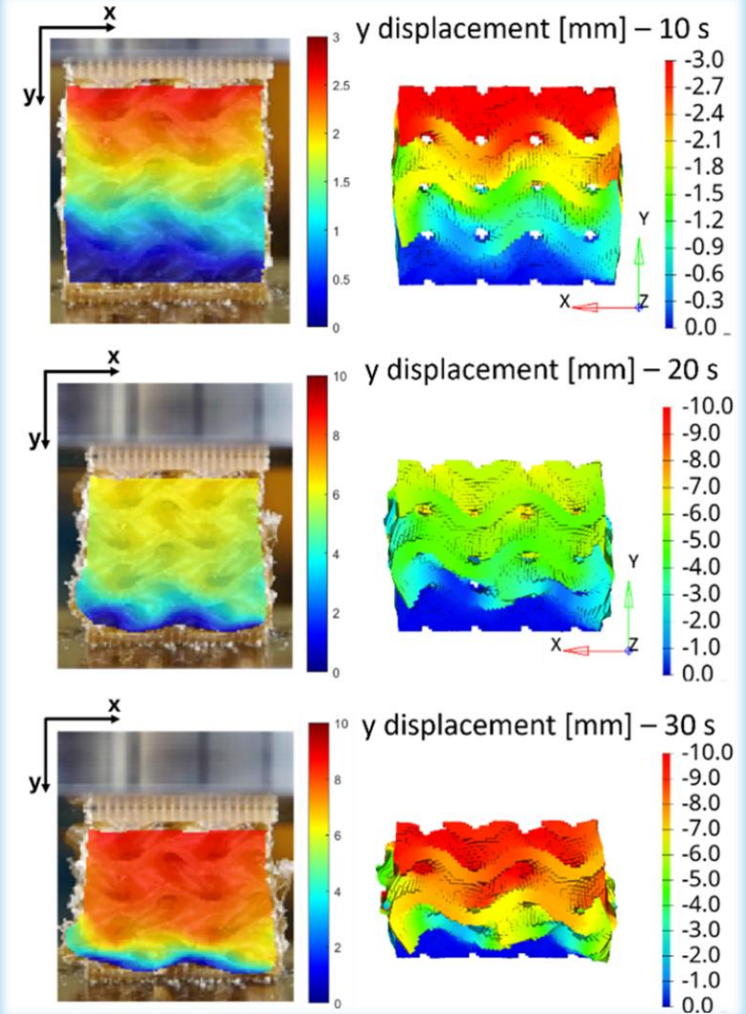
- **HyperMesh:**
 - gyroid lattice → 0.6 mm hexahedral mesh
 - plates → 1 mm quad elements mesh
- **Contacts friction coefficients:**
 - 0.50 for lattice–plates couple
 - 0.45 for lattice self-contact
- **Material laws:**
 - Johnson–Cook for PEKK
 - linear-elastic for steel plates



| | E [GPa] | ν | P [kg/m ³] | σ_{MAX} [MPa] | ϵ_f | ϵ_{dam} |
|--------------|---------|-------|------------------------|----------------------|--------------|------------------|
| PEKK | 2.3 | 0.4 | 1280 | 64 | 0.3127 | 0.2536 |
| Steel plates | 210 | 0.3 | 7850 | / | / | / |



Validated FEM model as a foundation for future design of PEKK-based porous orthopedic devices



Tests on acetabular cups

Short-term deformation under diametral compression



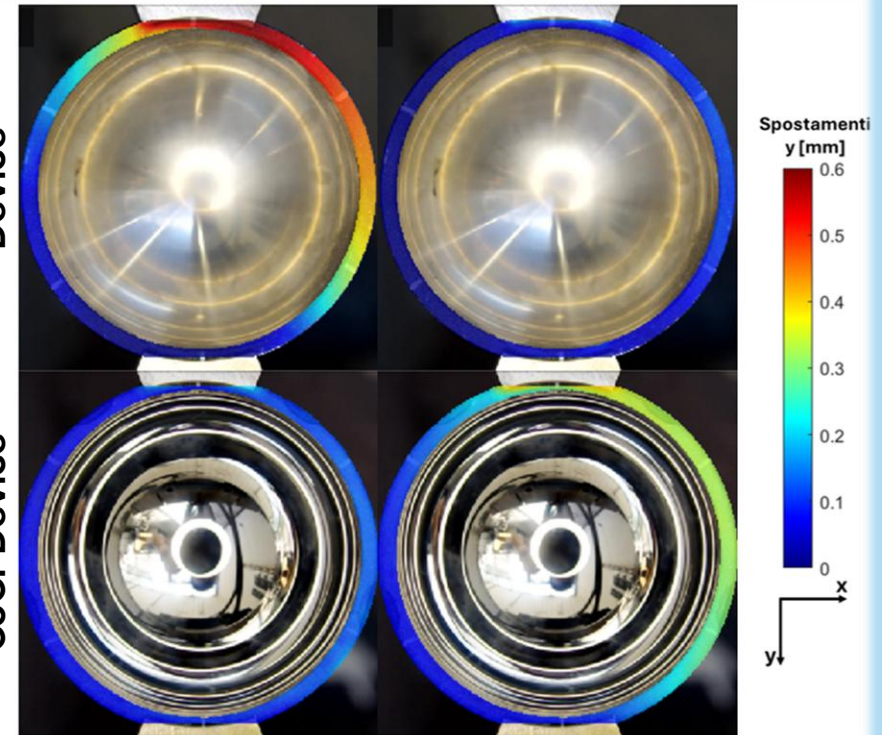
- press-fit acetabular shells
- Ti6Al4V ELI alloy by EBM
- external porous region with rhombic dodecahedral lattice (3 mm unit cell)

Tests

- 1000 N diametrical two-point compressive load (120° rotation)
- internal diameter measurements before, during and after loading

MTORTHO
Device

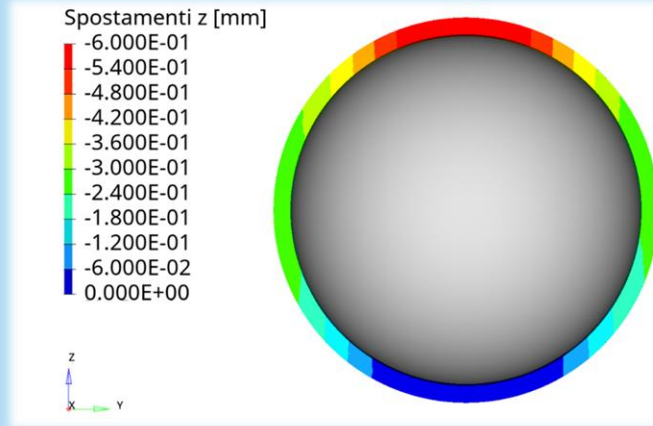
Standard
CoCr Device



Introduction of porosity did not compromise short-term mechanical integrity of the implant

Tests on acetabular cups

Numerical modelling and FEM validation

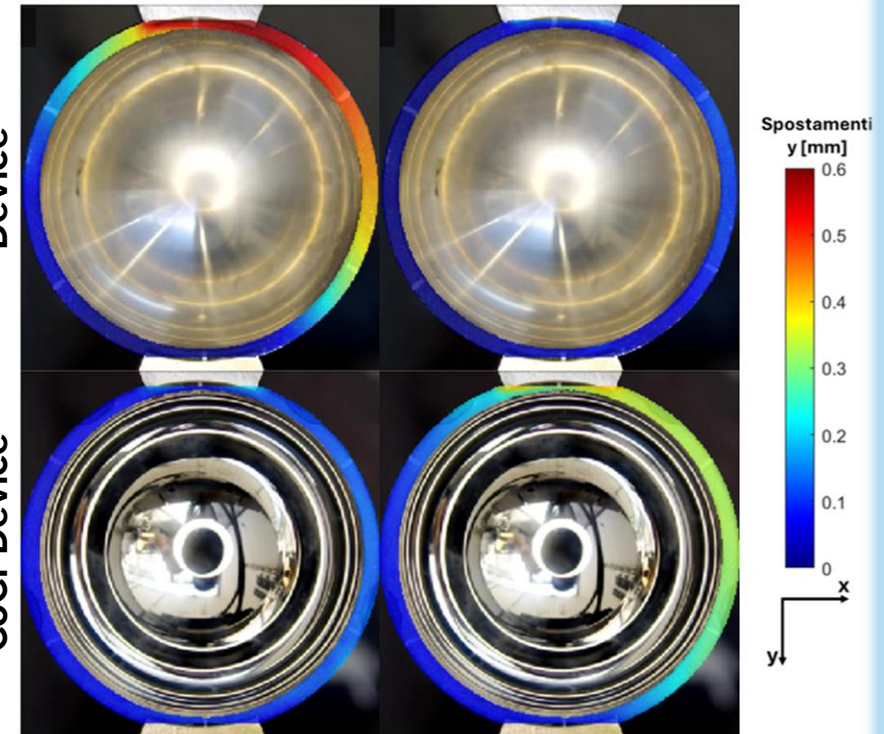


- 0.5 mm CTETRA first order elements
- equivalent isotropic material for net
- linear static

| | E [GPa] | ν | σ_{MAX} [MPa] | σ_y [MPa] |
|----------------|---------|-------|----------------------|------------------|
| Ti6Al4V (melt) | 108 | 0.3 | 965 | 873 |
| RD 3 mm (net) | 0.224 | 0.3 | 11.1 | / |

MTORTHO
Device

Standard
CoCr Device

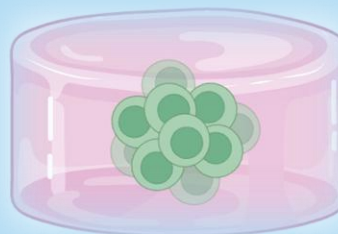


→ Validated FEM model for further parametric analyses on implant geometry and thickness

Immunomodulatory hydrogels

for bone tissue regeneration

- Bone healing is regulated by **immune** and **inflammatory processes**
- Excessive or **prolonged inflammation** impairs regeneration
- Timely immune regulation promotes **osteogenesis** and vascularization



Immunomodulatory cell-based strategies

Role of hydrogels

- enable localized delivery of therapeutic cells
- create immunomodulatory niches at the defect site

Regulatory T cells (Tregs)

- control excessive immune responses
- support tissue repair and immune homeostasis

Immunomodulatory hydrogels

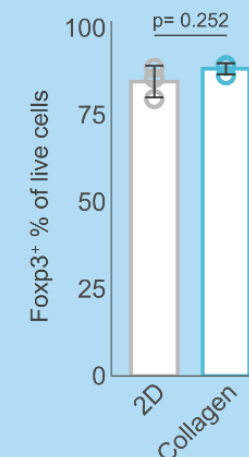
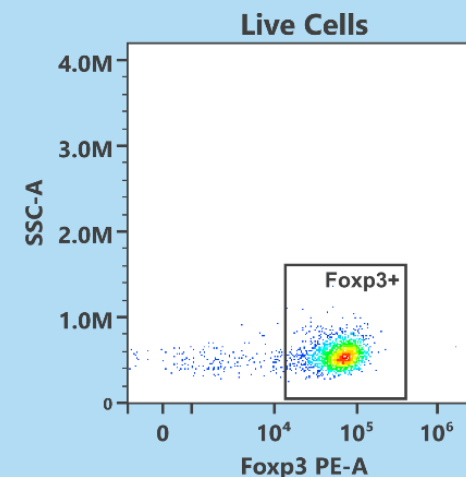
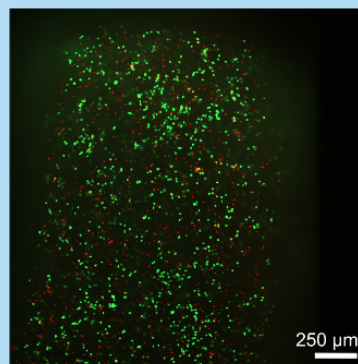
for bone tissue regeneration

Protocol and analysis:

- 1) CD4⁺ T cells isolation from mice spleens
- 2) Differentiation into Tregs with IL-2 and TGF- β for 4 days
- 3) Encapsulation in Collagen and GelMA hydrogels (500,000 cells/gel)
- 4) Live/Dead staining and flow cytometry for Foxp3 expression after 2 days

Results:

- High cell **viability**
- Homogeneous **distribution** of cells
- Preserved **Foxp3** expression after encapsulation



→ Toward GelMA hydrogels

Collagen: excellent bioactivity but fast degradation

GelMA: tunable stiffness, degradation, and stability

Thanks for your attention